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10/081,969	02/22/2002	Cheng Cheng	4-31704A/GTI	4496	
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DLA PIPER RUDNICK GRAY CARY U.S. LLP 1200 NINETEENTH STREET			MARVICH, MARIA		
			ART UNIT	PAPER NUMBER	
N.W. WASHIN	GTON, DC 20036-2412		1633		
			DATE MAILED: 09/19/2005	5	

Please find below and/or attached an Office communication concerning this application or proceeding.

		
	Application No.	Applicant(s)
	10/081,969	CHENG ET AL.
Office Action Summary	Examiner	Art Unit
	Maria B. Marvich, PhD	1633
The MAILING DATE of this communication Period for Reply	appears on the cover sheet wit	h the correspondence address
A SHORTENED STATUTORY PERIOD FOR RETHE MAILING DATE OF THIS COMMUNICATION - Extensions of time may be available under the provisions of 37 CF after SIX (6) MONTHS from the mailing date of this communication if the period for reply specified above is less than thirty (30) days, If NO period for reply is specified above, the maximum statutory period for reply within the set or extended period for reply will, by sany reply received by the Office later than three months after the rearned patent term adjustment. See 37 CFR 1.704(b).	ON. FR 1.136(a). In no event, however, may a re n. a reply within the statutory minimum of thirty eriod will apply and will expire SIX (6) MONT statute, cause the application to become ABA	ply be timely filed (30) days will be considered timely. (HS from the mailing date of this communication. ANDONED (35 U.S.C. § 133).
tatus		
1) Responsive to communication(s) filed on	02 February 2005.	
	This action is non-final.	
3) Since this application is in condition for all	owance except for formal matte	ers, prosecution as to the merits is
closed in accordance with the practice und		•
isposition of Claims		
4)⊠ Claim(s) <u>1-11,13,14,16-45,47-51,58,59 an</u>	d 63 is/are pending in the appli	cation.
4a) Of the above claim(s) is/are with		
5) Claim(s) is/are allowed.		
6) Claim(s) <u>1-11,13,14,16-45,47-51,58,59 and</u>	d 63 is/are rejected.	
7) Claim(s) is/are objected to.		
8) Claim(s) are subject to restriction as	nd/or election requirement.	
Application Papers	. ·	
9)☐ The specification is objected to by the Exar	miner.	
10) The drawing(s) filed on is/are: a)		ov the Examiner.
Applicant may not request that any objection to		
Replacement drawing sheet(s) including the co	rrection is required if the drawing(s	s) is objected to. See 37 CFR 1.121(d).
11) The oath or declaration is objected to by the	e Examiner. Note the attached	Office Action or form PTO-152.
riority under 35 U.S.C. § 119		
12) Acknowledgment is made of a claim for for	eign priority under 35 U.S.C. §	119(a)-(d) or (f).
a)□ All b)□ Some * c)□ None of:		
1. Certified copies of the priority docum		
2. Certified copies of the priority docum		
3. Copies of the certified copies of the		eceived in this National Stage
application from the International Bu		
* See the attached detailed Office action for a	list of the certified copies not re	eceived.
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ttachment(s)		•
Notice of References Cited (PTO-892)		Immary (PTO-413)
) 🔲 Notice of Draftsperson's Patent Drawing Review (PTO-948)		/Mail Date ormal Patent Application (PTO-152)
 Information Disclosure Statement(s) (PTO-1449 or PTO/SB Paper No(s)/Mail Date 	3/08) 5) Notice of Inf. 6) Other:	

Application/Control Number: 10/081,969

Art Unit: 1633

DETAILED ACTION

This office action is in response to an amendment filed 7/15/05. Claims 12, 15, 46, 52-57, 60-61 and 65-66 have been cancelled. Claims 1, 7, 23 and 40 have been amended. Claims 1-11, 13, 14, 16-45, 47-51, 58, 59 and 63 are pending in the instant application.

Response to Amendment

Any rejection of record in the previous action not addressed in this office action is withdrawn. There are no new grounds of rejection herein and therefore, this action is final.

Claim Rejections - 35 USC § 102

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless -

(e) the invention was described in (1) an application for patent, published under section 122(b), by another filed in the United States before the invention by the applicant for patent or (2) a patent granted on an application for patent by another filed in the United States before the invention by the applicant for patent, except that an international application filed under the treaty defined in section 351(a) shall have the effects for purposes of this subsection of an application filed in the United States only if the international application designated the United States and was published under Article 21(2) of such treaty in the English language.

Claims 1, 3-6, 8, 9, 14, 16, 18-25, 29, 35-38, 40, 48, 58 and 59 are rejected under 35 U.S.C. 102(e) as being anticipated by Johnson et al (US 2004/0151696; see entire document). This rejection is maintained for reasons of record in the office action mailed 4/20/05 and restated below. The rejection has been extended to newly amended claim 40.

Johnson et al teach an oncolytic vectors and particles that comprise in sequential order in a 5' to 3' direction, a left ITR, termination signals from the inserted transgene (CD) which has

Application/Control Number: 10/081,969

Art Unit: 1633

been isolated from its genetic source and inserted into the viral vector, a human E2F-1 promoter driving expression of E4 gene and a right ITR as recited in claim 1 (see e.g. figure 4, example 1 and paragraph 60). Furthermore, the vector is based upon Ad5 as recited in claim 40 (see e.g. paragraph 94 or 96). The E2F-1 promoter can be considered to be tissue specific as it is specific for Rb defective cancer cells thus in the case that E2F is linked to E1A, the E2F promoter driving expression of E4 is a tissue specific promoter (see e.g. paragraph 32). Thus the vector is designed to replicate in Rb defective cells but not in normal cells (see e.g. paragraph 88). The CR region can be deleted from the E1A gene, which deletes sequences upstream of the termination signal (see e.g. paragraph 88-91). According to the examples, the E3 19k gene is deleted and the cytosine deaminase is inserted into the vector (see e.g. example 7). Otherwise the E3 gene is present (paragraph 110 and figure 4b). However, the gene can be inserted into the E1B region, which would under most conditions inactivate the E1B region (see e.g. paragraph 0101). The vector further comprises a coding sequence such as for cytokines such as TNF α or IFNy (an immunomodulatory protein as well as antigenic protein), MIP3, cell suicide or apoptosis inducing proteins such as thymidine kinase, which can be inserted into the E3 19K or 14.7 K region (see e.g. paragraphs 101). Furthermore, tumor associated antigens or polypeptides that bind to receptors present on the tumor cell are contemplated (see e.g. paragraph 101).

Page 3

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person

having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.

Claims 1-9, 14, 16, 18-25, 29, 35-38, 40, 41, 43, 48, 58, 59, 62, 63, 64, 67, 69-76, 78, 82 and 83 are rejected under 35 U.S.C. 103(a) as being unpatentable over Johnson et al (US 2004/0151696; see entire document) in view of George and Blazing (5,880,102; see entire document) as evidenced by Hearing and Shenk (Cell, 1983, pages 695-703; see entire document). This rejection is maintained for reasons of record in the office action mailed 4/20/05 and restated below.

Applicants claim a recombinant viral vector comprising in sequential order a left ITR, a termination signal, an E2F responsive promoter operably linked to a gene required for replication, a packaging signal and a right ITR.

Johnson et al teach an oncolytic vectors and particles that comprise in sequential order in a 5' to 3' direction, a left ITR, termination signals from the inserted transgene (CD) which has been isolated from its genetic source and inserted into the viral vector, a human E2F-1 promoter driving expression of E4 gene and a right ITR as recited in claim 1 (see e.g. figure 4, example 1 and paragraph 60). Furthermore, the vector is based upon Ad5 as recited in claim 40 (see e.g. paragraph 94 or 96). The E2F-1 promoter can be considered to be tissue specific as it is specific for Rb defective cancer cells thus in the case that E2F is linked to E1A, the E2F promoter driving expression of E4 is a tissue specific promoter (see e.g. paragraph 32). Thus the vector is designed to replicate in Rb defective cells but not in normal cells (see e.g. paragraph 88). The CR region can be deleted from the E1A gene, which deletes sequences upstream of the termination signal (see e.g. paragraph 88-91). According to the examples, the E3 19k gene is deleted and the cytosine deaminase is inserted into the vector (see e.g. example 7). Otherwise

the E3 gene is present (paragraph 110 and figure 4b). However, the gene can be inserted into the E1B region, which would under most conditions inactivate the E1B region (see e.g. paragraph 0101). The vector further comprises a coding sequence such as for cytokines such as TNFα or IFNγ (an immunomodulatory protein as well as antigenic protein), MIP3, cell suicide or apoptosis inducing proteins such as thymidine kinase, which can be inserted into the E3 19K or 14.7 K region (see e.g. paragraphs 101). Furthermore, tumor associated antigens or polypeptides that bind to receptors present on the tumor cell are contemplated (see e.g. paragraph 101). The adenovirus can be altered for selective delivery to neoplastic cells based upon a cell surface protein, which binds an immunoglobin or immunoliposomes (see e.g. paragraph 0106).

Johnson et al do not teach either that the termination signal is a SV40 polyadenylation sequence or that this sequence is positioned 5' to the E1a gene or that the packaging signal is positioned 3' of the gene required for replication and prior to the right ITR.

George and Blazing teach generation of an adenovirus vector, i.e. Ad Pac-Bgal in which a SV40 polyadenylation sequence is 3' to the left ITR and 5' of E1A gene (see e.g. figure 47). Placement of the SV40 polyadenylation sequences functions to terminate expression of sequences that are 3' to the left ITR. Furthermore, the e1a enhancer and packaging sequences are inserted at the 3' end of the vector (see e.g. figure 47 and col 2, line 6-10). In this manner, the E1A enhancer regions, which are intertwined with the packaging signal are non-operative as evidenced by Hearing and Shenk while the packaging functions are retained (see e.g. abstract). Therefore, in the vector of George and Blazing expression of E1A or gene substituted for E1A can be expressed by for example tissue-specific promoters without read through from the E1A

enhancer. The results of these manipulations are deletion of the region of Ad5 that comprises nucleotides 103-551.

It would have been obvious to one of ordinary skill in the art at the time the invention was made to move the packaging signal in the vector taught by Johnson et al to the 3' end of the vector as taught by George and Blazing as evidenced by Hearing and Shenk because Johnson et al teach that it is within the ordinary skill of the art to generate recombinant virus in which the E4 gene is under control of the E2F promoter and because George and Blazing teach that it is within the ordinary skill of the art to move the packaging signal to the 3' end of the adenovirus. One would have been motivated to do so in order to receive the expected benefit of reducing E1A in the cell such that the virus is E1a-RB replication deficient as described by Johnson et al (see e.g. paragraph 0089). Based upon the teachings of the cited references, the high skill of one of ordinary skill in the art, and absent evidence to the contrary, there would have been a reasonable expectation of success to result in the claimed invention.

Claims 49-51 are rejected under 35 U.S.C. 103(a) as being unpatentable over Johnson et al (US 2004/0151696; see entire document) in view of George and Blazing (5,880,102; see entire document) as evidenced by Hearing and Shenk (Cell, 1983, pages 695-703; see entire document) further in view of Krasnykh et al (JVI, 1998, pages 1844-1852). This rejection is maintained for reasons of record in the office action mailed 4/20/05 and restated below.

Applicants claim a recombinant viral vector comprising in sequential order a left ITR, a termination signal, an E2F responsive promoter operably linked to a gene required for replication

a packaging signal and a right ITR. Particles comprise a targeting ligand in the HI loop of the fiber protein.

The teachings of Johnson and George and Blazing and Hearing and Shenk are as above except; none teach insertion of targeting ligands into the HI loop of the fiber protein.

Krasnykh et al teach targeted vectors that are capable of gene delivery to selected cell types in vivo by incorporation of heterologous ligands in the HI loop of the fiber protein. The HI loop possesses several properties, which predicts its utility as a site for ligand incorporation. Specifically, the HI loop does not contribute to the intramolecular interaction in the knob such that the ligand will not affect trimerization. The HI loop is exposed outside the knob and insertions should not affect correct folding of the entire knob (see e.g. page 1845, col 1). Krasnykh et al propose insertions into the HI loop for generation of retargeted vectors that can be directed to specific cells (see e.g. page 1851, col 2).

It would have been obvious to one of ordinary skill in the art at the time the invention was made to specifically insert the targeting ligands taught by Johnson et al into HI loop of the fiber protein as taught by Krasnykh et al because Johnson et al teach that it is within the ordinary skill of the art to generate recombinant virus in which a targeting ligand is used to specifically target cells and because Krasnykh et al teach that it is within the ordinary skill of the art to insert targeting ligands into the HI loop of the fiber protein. One would have been motivated to do so in order to receive the expected benefit of cell retargeting in which the ligand is exposed outside the knob and insertions should not affect correct folding of the entire knob or trimerization. Based upon the teachings of the cited references, the high skill of one of ordinary skill in the art,

and absent evidence to the contrary, there would have been a reasonable expectation of success to result in the claimed invention.

Claims 26-28 and 30-34 are rejected under 35 U.S.C. 103(a) as being unpatentable over Johnson et al (US 2004/0151696; see entire document) in view of George and Blazing (5,880,102; see entire document) as evidenced by Hearing and Shenk (Cell, 1983, pages 695-703; see entire document) further in view of Compagni et al (Cancer Research, 2000, pages 7163-7169; see entire document) or Griscelli et al (PNAS, 2000, pages 6698-6703; see entire document) or Kaplan et al (American Association of Immunologists, 1999, pages 699-707) or Ali et al (Gene Therapy, 1998, page 1561-1565). This rejection is maintained for reasons of record in the office action mailed 4/20/05 and restated below.

Applicants claim a recombinant viral vector comprising in sequential order a left ITR, a termination signal, an E2F responsive promoter operably linked to a gene required for replication a packaging signal and a right ITR. The vector expresses tumor-associated antigens such as gp100, MART-1 or trp or anti angiogenic proteins such as angiostatin or antagonists of FGF or inhibitors of PDGF or fragments of TrpRS or antibodies that block inhibitory signals.

The teachings of Johnson and George and Blazing and Hearing and Shenk are as above except; none of the teachings teach that the vector also expresses tumor-associated antigens such as gp100, MART-1 or trp or anti angiogenic proteins such as angiostatin or antagonists of FGF or inhibitors of PDGF or fragments of TrpRS or antibodies that block inhibitory signals.

Griscelli et al teach that angiostatin is a specific inhibitor of endothelial cell proliferation. Griscelli et al express angiostatin using adenovirus to antagonize growth of endothelial cells

Application/Control Number: 10/081,969

Art Unit: 1633

during the early stages of tumor angiogenesis (see e.g. page 6698, col 2, last paragraph and page 6703, col 1-2). Furthermore, in combination with cytotoxic therapies such as radiotherapy angiostatin enhances the response of larger tumors to angiostatin (see e.g. page 6698, col 2, paragraph 3).

Compagni et al teach use of soluble FGFR (sFGFR) as an antagonist of FGF function.

Adenovirus expressing sFGFR were used to specifically repress proliferation and differentiation induced by FGF1 (see e.g. page 7168, col 1, paragraph 2). sFGFR was potent in inhibiting tumor growth and impairing tumor angiogenesis (see e.g. abstract and page 7168, col 2, paragraph 1).

Kaplan et al teach that dendritic cells transduced with sequences encoding several tumor associated antigens or peptides induce an Ag specific CTL response resulting in protection from tumor challenge and resulting in regression of tumors (see e.g. abstract). The cells were transduced with adenovirus expressing MAA, gp100, tyrosinase related peptide (TRP) and MART-1.

Ali et al teach expression of CTLA4-Ig from adenovirus to inhibit destruction of transduced cells (see e.g. page 1563, bridging paragraph col 1-2). Expression of CTLA4-Ig block host immune responses to the adenovirus for enhanced transgene expression by blocking the B7-CD28 interactions between antigen presenting cells and T cells to prevent the costimulatory signals required for T cell survival and proliferation (see e.g., abstract).

Otani et al teach that a fragment of human TrpRS functions as a potent antagonist of VEGF angiogenesis (see e.g. abstract).

Zhao et al teach adenovirus mediated decorin gene transfer to prevent TGF-\beta induced inhibition of lung morphogenesis (see e.g. abstract). Decorin was expressed using a recombinant

adenovirus, which ameliorated excessive levels of TGF-β signaling in the developing lung. Excessive TGF-b results in abnormalities of lung growth, differentiation and development (see e.g. L413, col 1, paragraph 2).

It would have been obvious to one of ordinary skill in the art at the time the invention was made to use the viral vector taught by Johnson et al in view of George and Blazing as evidenced by Hearing and Shenk to express the transgenes taught by Griscelli et al, Compagni et al, Kaplan et al, Ali et al, Otani et al and Zhao et al because Johnson et al in view of George and Blazing as evidenced by Hearing and Shenk teach that it is within the ordinary skill of the art to generate recombinant adenovirus that express heterologous genes and Griscelli et al, Compagni et al, Kaplan et al, Ali et al, Otani et al and Zhao et al teach that it is within the ordinary skill of the art to express transgenes that express products such as angiostatin, decorin, fragments of TrpRS, CTLA4-Ig MAA, gp100, tyrosinase related peptide (TRP) and MART-1 and sFGFR. One would have been motivated to do so in order to receive the expected benefit of antagonizing growth of endothelial cells during the early stages of tumor angiogenesis (see e.g. Griscelli et al) or to inhibit tumor growth and impair tumor angiogenesis (see Compagni et al and Otani et al) or for protection from tumor challenge and for regression of tumors (see e.g. Kaplan et al) or for enhanced transgene expression (see Ali et al) or to prevent TGF-\beta induced inhibition of lung morphogenesis (see e.g. Zhao et al) and Johnson et al teach viral vector to be used to treat cancer. Based upon the teachings of the cited references, the high skill of one of ordinary skill in the art, and absent evidence to the contrary, there would have been a reasonable expectation of success to result in the claimed invention.

Response to Argument

Applicants traverse the claim rejections under 35 U.S.C. 102 and 103 on pages 11-15 of the amendment filed 7/15/05. Applicants argue that Johnson does not anticipate the claims as amended and George and Blazing and Hearing and Shenk do not cure the teachings of Johnson et al.

Applicants' arguments filed 7/15/05 have been fully considered but they are not persuasive. Applicants appear to argue that the amendment that the termination signal is isolated form its genetic source distinguishes the instant invention from the prior art. However, according to Johnson et al, the CD gene is isolated from it source and inserted into the vector. Therefore, the termination signal that is associated with the gene is also inserted into the vector. This termination signal is 5' of the E2F promoter and the gene essential for replication. Thus, Johnson et al does anticipate the instant invention. Hearing and Shenk and George and Blazing do not need to teach what the primary reference teaches.

Conclusion

Claims 1-9, 14, 16, 18-38, 40, 41, 43, 48-51, 58, 59 and 63 are rejected.

Claims 10, 11, 13, 17, 39, 42, 44, 45 and 47 are objected to as being dependent upon a rejected base claim, but would be allowable if rewritten in independent form including all of the limitations of the base claim and any intervening claims.

THIS ACTION IS MADE FINAL. Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the mailing date of this final action.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Maria B. Marvich, PhD whose telephone number is (571)-272-0774. The examiner can normally be reached on M-F (6:30-3:00).

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, David Nguyen, PhD can be reached on (571)-272-0731. The Central Fax number for official documents is (703)-872-9305. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

Maria B Marvich, PhD Examiner Art Unit 1633

September 08, 2005

JAMES KETTER
PRIMARY EXAMINER